

Commentary

WRN Protein and Werner Syndrome

Jianyuan Luo, PhD

Abstract

Werner syndrome is an autosomal recessive disorder associated with premature aging and cancer predisposition. Cells from Werner syndrome patients show increased genomic instability and are hypersensitive to DNA damage agents. Werner syndrome is caused by mutations of the *WRN* gene. WRN protein is a member of RecQ DNA helicase family. It not only contains a conserved 3'-5' helicase domain as other members of the RecQ family but also contains a unique 3'-5' exonuclease domain. WRN recognizes specific DNA structures as substrates which are intermediates of DNA metabolism. WRN interacts with many other proteins, which function in telomere maintenance, DNA replication, and DNA repair through different pathways.

[N A J Med Sci. 2010;3(4):205-207.]

Werner Syndrome and its Cellular Phenotype

Werner syndrome is a human autosomal recessive disorder that displays symptoms of premature aging, including early graying and thinning of hair, wrinkling and ulceration of skin, atherosclerosis, osteoporosis, and cataracts. In addition, Werner syndrome patients exhibit an increased incidence of diabetes mellitus type 2, hypertension, and are highly disposed to the emergence of benign and malignant neoplasms.¹ Werner syndrome caused by mutation of the *WRN* gene, a member of the RecQ DNA helicase family.² There are five known human RecQ helicases.³ Mutations in two other family members, BLM and RecQ4, are responsible for the two other premature aging syndromes, Bloom's⁴ and Rothmund Thomson,⁵ respectively. The *WRN* gene encodes a 1,432-amino acid protein² that contains both 3'→5' helicase and 3'→5' exonuclease activities.⁶⁻⁹

The cells from patients with Werner syndrome show premature replicative senescence compared with cells derived from normal individuals.¹⁰ The Werner syndrome cellular phenotype suggests correlations among faulty DNA

metabolism, genomic instability, and senescence. Werner syndrome cells show hypersensitivity to selected DNA-damaging agents including 4-nitroquinoline-1-oxide (4NQO),¹¹ topoisomerase inhibitors,¹² and certain DNA cross-linking agents.¹³ Comparing with normal cells, Werner syndrome cells also exhibit increased genomic instability including higher levels of DNA deletions, translocations, and chromosomal breaks.^{14,15} These studies suggest that WRN plays an important role in DNA repair, replication, and recombination pathways.

Werner Syndrome Mouse Model

Two knockout mouse models for Werner syndrome have been established by using different strategies, and displayed a distinct phenotype. Lebel and Leder targeted the exons that encode motifs III and IV of the helicase domain of mouse WRN. The targeted gene expresses a fully translated WRN protein that lacks 121 amino acids of the helicase domain. Therefore, it is expected that it lacks helicase activity but retains other putative functions of the WRN protein. The homozygous mice were born viable, and during the first year, the mice appeared normal. By 24 months, 62% of the homozygous mice had developed some kind of hyperplasia or tumor.¹² WRN^{Δhel/Δhel} p53^{-/-} double-knockout mice were shown to develop tumors more rapidly than the p53^{-/-} parental line. Indeed, by 4–5 months, 50% of the double-knockout population had developed tumors.¹⁶ The other knockout mouse, generated by Lombard and co-workers,¹⁷ targeted the last exon of the helicase domain, which results in the expression of a truncated protein, a situation similar to that seen in most human Werner syndrome cases. Moreover, as is seen with many of the WS mutations in humans, these authors could not detect expression of the truncated protein. This mouse allele, therefore, can be considered as WRN⁻.¹⁷ The WRN^{-/-} homozygous mutant mice were reported to be perfectly healthy. Also, WRN^{-/-} embryonic fibroblasts from these animals showed no signs of sensitivity to 4NQO or camptothecin, and there was no significant decrease in the replicative lifespan of these fibroblasts. However, p53^{-/-} WRN^{-/-} double-knockout mice died earlier, and the lack of p53 accelerated the mortality of WRN^{-/-} or WRN^{-/-} mice. The lack of an obvious phenotype in this WRN^{-/-} mice might be explained by the fact that telomerase is expressed constitutively in rodent cells, effectively masking any effect of a loss of WRN function.¹⁷ The finding of the mice with WRN and Terc double knock out exhibiting the typical Werner syndrome phenotype indicates that WRN regulating telomere function.^{18,19}

Received 10/14/2010; Revised 10/23/2010; Accepted 10/24/2010

Jianyuan Luo, PhD

Department of Medical & Research Technology

Department of Pathology

School of Medicine, University of Maryland

AHB 405A, 100 Penn Street, Baltimore, MD 21201

jluo@som.umaryland.edu

Biochemical Characteristics of the WRN Protein

The lack of phenotypes of WRN knockout mice underscores the need to study human WRN functions at biochemical and cellular level. WRN is a bipartite and bifunctional enzyme. In addition to the ATP-dependent 3'→5' helicase and DNA-dependent ATPase activities, it also possesses a functional 3'→5' exonuclease domain, which is similar to the exonuclease domain of *E. coli* DNA polymerase I.²⁰ The two enzymatic functions are separable from each other. The mutation WRND82A and E84A, disables exonuclease activity, but retains its helicase activity.⁷ Similarly, the ATPase and helicase activities are diminished when the Lys577 residue in the ATPase domain is mutated, but the exonuclease activity is not affected by this substitution.^{7,21} The helicase and exonuclease activities are physically separable, recombinant N-terminal fragments WRN1–368,²² and the minimal exonuclease domain WRN70–240²³ display exonuclease activity without helicase activity. Similarly, N terminal deletion derivatives, which lack the exonuclease domain, retain helicase activity. Although the two enzymatic activities are not affect each other, full function and regulation of catalytic activities require the presence of other regions of the protein, which could modify the activity of the minimal enzymatic domains.²⁴

The helicase activity of WRN shows very poor processivity as it preferentially unwound bubble substrates, forked structures and G-quadruplex DNA.²⁵ WRN was also capable of branch-migrating Holliday junctions over distances as great as 2.7 kb.²⁶ The 3'→5' exonuclease activity also displays low processivity. Exonucleolytic cleavage results in the production of 5'-deoxymonophosphates.⁸ It prefers 5' overhangs, bubbles, loops and Holliday junctions structures, but has little or no activity on double-stranded duplexes with blunt ends, partial duplexes with 3' overhangs, or single-strand DNA.²⁴

Functions of WRN Protein and its Subcellular Localization

The multi-functional nature of WRN, as well as the range of different symptoms seen in Werner syndrome patients, suggests that WRN may be a versatile enzyme with an involvement in diverse cellular processes. The finding that WRN interacts physically and functionally with other proteins required for DNA metabolism supports this notion. WRN was found to interact directly with numerous proteins, including proliferating cell nuclear antigen and Topoisomerase I,²⁷ Replication Protein A (RPA),²⁸ p53,^{29,30} the Ku complex,^{31,32} and DNA Pol δ.³³ In this regard, WRN may be a functional component of various cellular processes, including DNA replication, transcription, recombination, and repair.

WRN protein shows dynamic translocalization within the nucleus under different conditions. The WRN protein localizes to the nucleoli in a variety of cell types,³⁴ and this localization is modulated by DNA damage during cell cycle. Upon serum starvation or treatment with hydroxyurea (HU),

aphidicolin, 4NQO, etoposide or camptothecin, WRN migrates from nucleoli to discrete nuclear foci.^{26,30,35-37} The fact that DNA damage also induces the formation of RPA and RAD51 foci, and these co-localize with WRN almost fully (RPA), or partially (RAD51),³⁷ emphasizes the potential role of the WRN protein in DNA replication and DNA repair.

Conclusion Mark

The increasing number of WRN interacting proteins involving DNA replication, recombination and repair provide the strong evidence that WRN functions in multiple DNA metabolic processes. WRN participates in several DNA repair pathways including double strand break repair for both homologous recombination (HR) and non-homologous end-joining (NHEJ) pathways and base excision repair (BER) pathway. While it may not be essential in any individual process, WRN appears to have significant functional roles in these pathways. WRN resolves DNA intermediates that arise normally as a result of DNA repair and replication processes. However, the precise role of WRN protein in these processes still remains unclear. How the mutations of WRN protein contribute to the plethora of premature aging symptoms in Werner Syndrome is yet to be ascertained. Further study to address these important questions should lead to new insights regarding aging process.

References

1. Martin GM, Austad SN, Johnson TE. Genetic analysis of ageing: role of oxidative damage and environmental stresses. *Nat Genet.* 1996; 13(1):25-34.
2. Yu CE, Oshima J, Fu YH, et al. Positional cloning of the Werner's syndrome gene. *Science* 1996; 272(5259):258-262.
3. Hickson ID. RecQ helicases: caretakers of the genome. *Nat Rev Cancer.* 2003;3(3):169-178.
4. Ellis NA, Groden J, Ye TZ, et al. The Bloom's syndrome gene product is homologous to RecQ helicases. *Cell.* 1995;83(4):655-666.
5. Kitao S, Shimamoto A, Goto M, et al. Mutations in RECQL4 cause a subset of cases of Rothmund-Thomson syndrome. *Nat Genet.* 1999;22(1):82-84.
6. Gray MD, Shen JC, Kamath-Loeb AS, et al. The Werner syndrome protein is a DNA helicase. *Nat Genet.* 1997;17(1):100-103.
7. Huang S, Li B, Gray MD, Oshima J, Mian IS, Campisi J. The premature ageing syndrome protein, WRN, is a 3'→5' exonuclease. *Nat Genet.* 1998;20(2):114-116.
8. Kamath-Loeb AS, Shen JC, Loeb LA, Fry M. Werner syndrome protein: II. Characterization of the integral 3'→5' DNA exonuclease. *J Biol Chem.* 1998;273(51):34145-34150.
9. Orren DK, Brosh RM Jr, Nehlin JO, Machwe A, Gray MD, Bohr VA. Enzymatic and DNA binding properties of purified WRN protein: high affinity binding to single-stranded DNA but not to DNA damage induced by 4NQO. *Nucleic Acids Res.* 1999;27(17):3557-3566.
10. Martin GM, Sprague CA, Epstein CJ. Replicative life-span of cultivated human cells. Effects of donor's age, tissue, and genotype. *Lab Invest.* 1970;23(1):86-92.
11. Ogburn CE, Oshima J, Poot M, et al. An apoptosis-inducing genotoxin differentiates heterozygotic carriers for Werner helicase mutations from wild-type and homozygous mutants. *Hum Genet.* 1997;101(2):121-125.
12. Lebel M, Leder P. A deletion within the murine Werner syndrome helicase induces sensitivity to inhibitors of topoisomerase and loss of cellular proliferative capacity. *Proc Natl Acad Sci U S A.* 1998;95(22):13097-13102.
13. Poot M, Yom JS, Whang SH, Kato JT, Gollahon KA, Rabinovitch PS. Werner syndrome cells are sensitive to DNA cross-linking drugs. *FASEB J.* 2001;15(7):1224-1226.
14. Fukuchi K, Martin GM, Monnat RJ. Mutator phenotype of Werner syndrome is characterized by extensive deletions. *Proc Natl Acad Sci U S A.* 1989;86(15):5893-5897.

15. Stefanini M, Scappaticci S, Lagomarsini P, Borroni G, Berardesca E, Nuzzo F. Chromosome instability in lymphocytes from a patient with Werner's syndrome is not associated with DNA repair defects. *Mutat Res.* 1989;219(3):179-185.
16. Lebel M, Cardiff RD, Leder P. Tumorigenic effect of nonfunctional p53 or p21 in mice mutant in the Werner syndrome helicase. *Cancer Res.* 2001;61(5):1816-1819.
17. Lombard DB, Beard C, Johnson B, et al. Mutations in the WRN gene in mice accelerate mortality in a p53-null background. *Mol Cell Biol.* 2000;20(9):3286-3291.
18. Chang S, Multani AS, Cabrera NG, et al. Essential role of limiting telomeres in the pathogenesis of Werner syndrome. *Nat Genet.* 2004;36(8):877-882.
19. Laud PR, Multani AS, Bailey SM, et al. Elevated telomere-telomere recombination in WRN-deficient, telomere dysfunctional cells promotes escape from senescence and engagement of the ALT pathway. *Genes Dev.* 2005;19(21):2560-2570.
20. Mushegian AR, Bassett DE Jr, Boguski MS, Bork P, Koonin EV. Positionally cloned human disease genes: patterns of evolutionary conservation and functional motifs. *Proc Natl Acad Sci U S A.* 1997;94(11):5831-5836.
21. Shen JC, Gray MD, Oshima J, Kamath-Loeb AS, Fry M, Loeb LA. Werner syndrome protein. I. DNA helicase and DNA exonuclease reside on the same polypeptide. *J Biol Chem.* 1998;273(51):34139-34144.
22. Machwe A, Xiao L, Theodore S, Orren DK. DNase I footprinting and enhanced exonuclease function of the bipartite Werner syndrome protein (WRN) bound to partially melted duplex DNA. *J Biol Chem.* 2002;277(6):4492-4504.
23. Xue Y, Ratcliff GC, Wang H, et al. A minimal exonuclease domain of WRN forms a hexamer on DNA and possesses both 3'-5' exonuclease and 5'-protruding strand endonuclease activities. *Biochemistry.* 2002;41(9):2901-2912.
24. Bachrati CZ, Hickson ID. RecQ helicases: suppressors of tumorigenesis and premature aging. *Biochem J.* 2003;374(Pt 3):577-606.
25. Mohaghegh P, Karow JK, Brosh RM Jr, Bohr VA, Hickson ID. The Bloom's and Werner's syndrome proteins are DNA structure-specific helicases. *Nucleic Acids Res.* 2001;29(13):2843-2849.
26. Constantinou A, Tarsounas M, Karow JK, et al. Werner's syndrome protein (WRN) migrates Holliday junctions and co-localizes with RPA upon replication arrest. *EMBO Rep.* 2000;1(1):80-84.
27. Lebel M, Spillare EA, Harris CC, Leder P. The Werner syndrome gene product co-purifies with the DNA replication complex and interacts with PCNA and topoisomerase I. *J Biol Chem.* 1999;274(53):37795-37799.
28. Brosh RM Jr, Orren DK, Nehlin JO, et al. Functional and physical interaction between WRN helicase and human replication protein A. *J Biol Chem.* 1999;274(26):18341-18350.
29. Blander G, Kipnis J, Leal JF, Yu CE, Shellenberg GD, Oren M. Physical and functional interaction between p53 and the Werner's syndrome protein. *J Biol Chem.* 1999;274(41):29463-29469.
30. Brosh RM, Jr, Karmakar P, Sommers JA, et al. p53 modulates the exonuclease activity of Werner syndrome protein. *J Biol Chem.* 2001;276(37):35093-35102.
31. Cooper MP, Machwe A, Orren DK, Brosh RM Jr, Ramsden DA, Bohr VA. Ku complex interacts with and stimulates the Werner protein. *Genes Dev.* 2000;14(8):907-912.
32. Orren DK, Machwe A, Karmakar P, Piotrowski J, Cooper MP, Bohr VA. A functional interaction of Ku with Werner exonuclease facilitates digestion of damaged DNA. *Nucleic Acids Res.* 2001;29(9):1926-1934.
33. Szekely AM, Chen YH, Zhang C, Oshima J, Weissman SM. Werner protein recruits DNA polymerase delta to the nucleolus. *Proc Natl Acad Sci U S A.* 2000;97(21):11365-11370.
34. Marciniak RA, Lombard DB, Johnson FB, Guarente L. Nucleolar localization of the Werner syndrome protein in human cells. *Proc Natl Acad Sci U S A.* 1998;95(12):6887-6892.
35. Nguyen DT, Rovira I, Finkel T. Regulation of the Werner helicase through a direct interaction with a subunit of protein kinase A. *FEBS Lett.* 2002;521(1-3):170-174.
36. Gray MD, Wang L, Youssoufian H, Martin GM, Oshima J. Werner helicase is localized to transcriptionally active nucleoli of cycling cells. *Exp Cell Res.* 1998;242(2):487-494.
37. Sakamoto S, Nishikawa K, Heo SJ, Goto M, Furuichi Y, Shimamoto A. Werner helicase relocates into nuclear foci in response to DNA damaging agents and co-localizes with RPA and Rad51. *Genes Cells.* 2001;6(5):421-430.